

Determining Possible Fragments Based On Experimental Restrictions

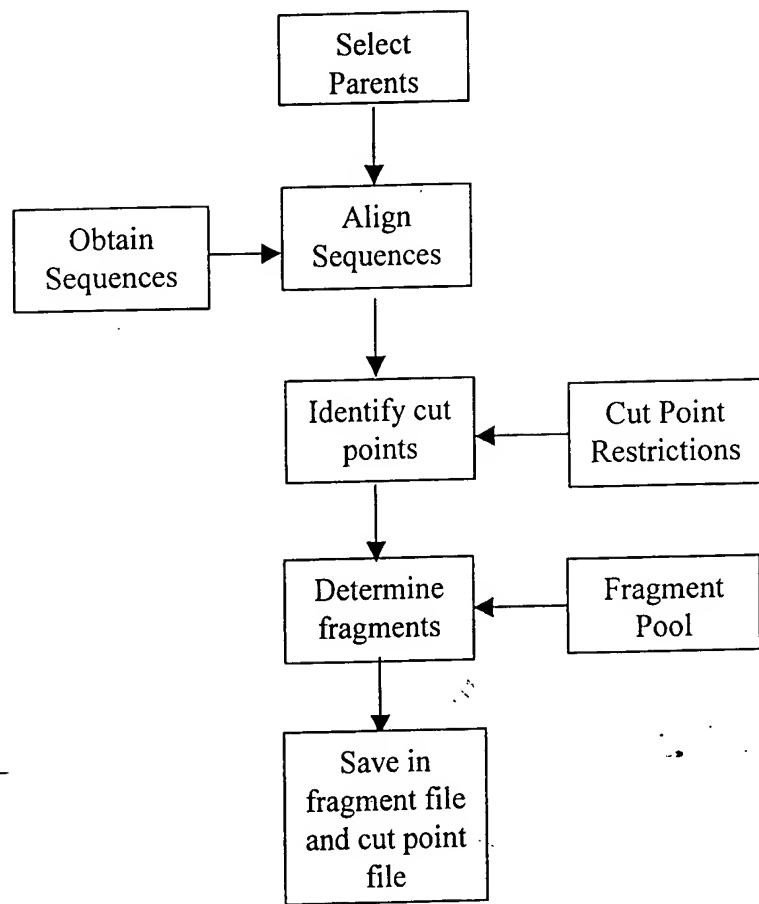


FIG. 1A

Determining the Schema Disruption Profile for a Structure

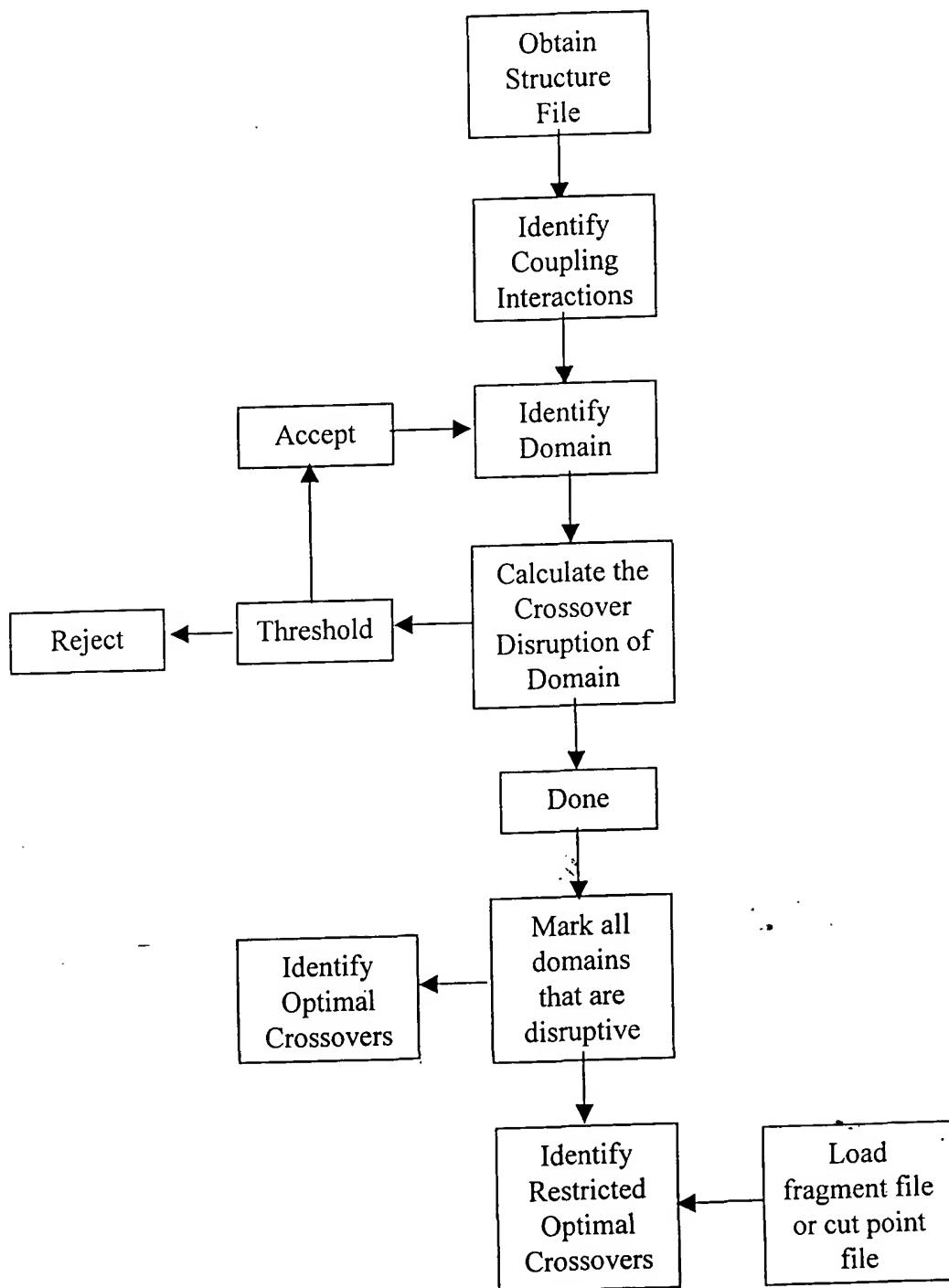


FIG. 1B

1 2 3 4 5 6 7 8 9 10 11 12

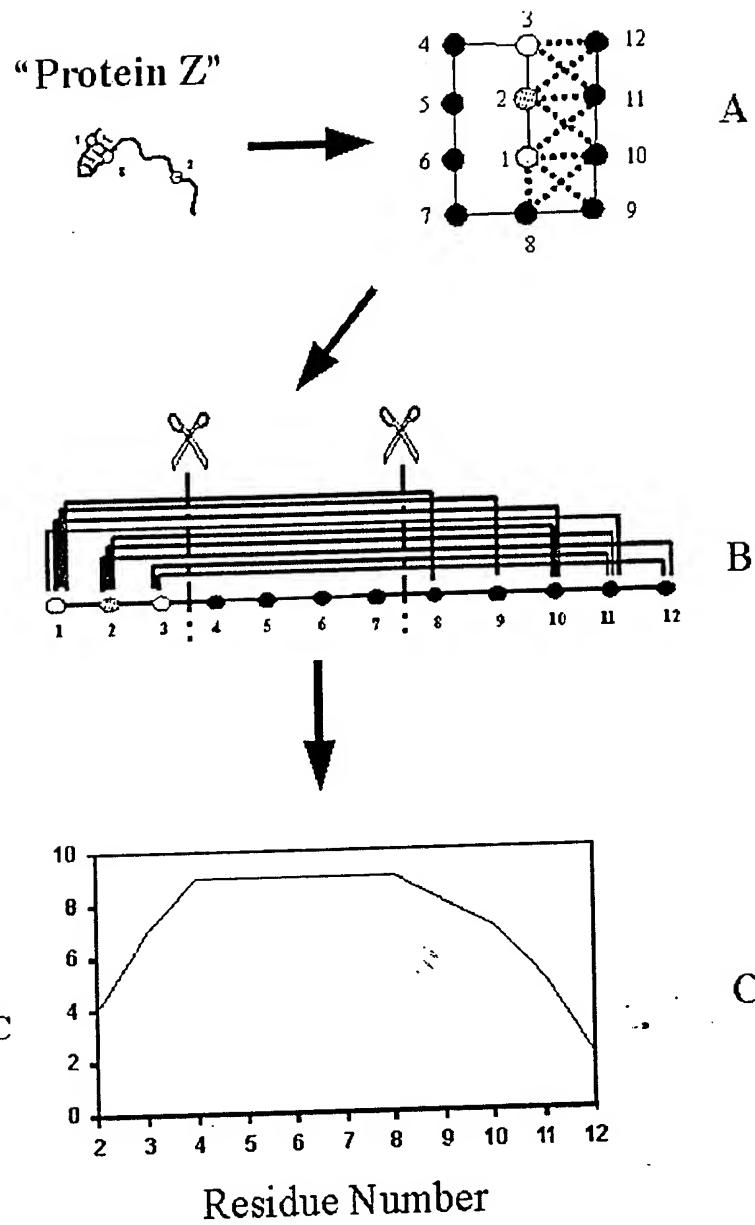


FIG. 2

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FIG. 3

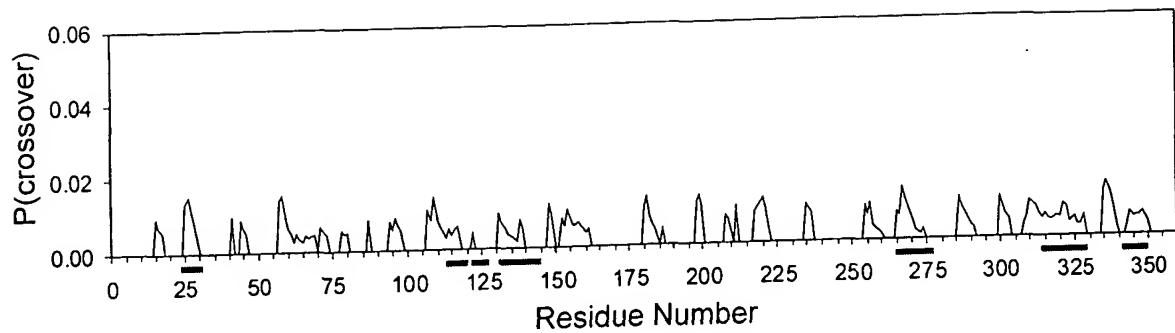


FIG. 4A

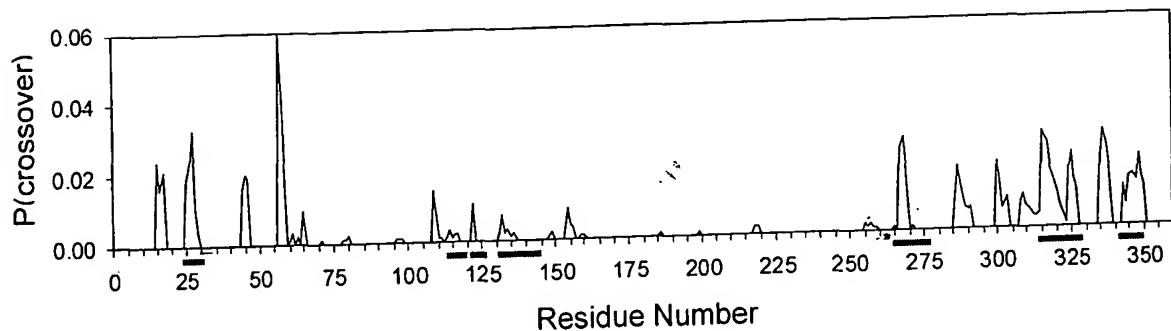


FIG. 4B

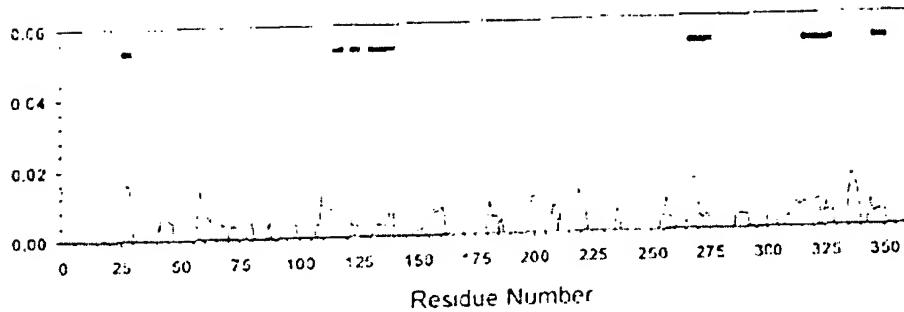


FIG. 4C

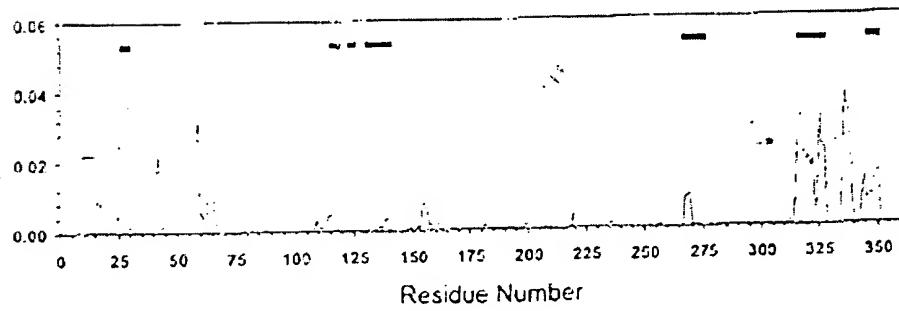


FIG. 4D

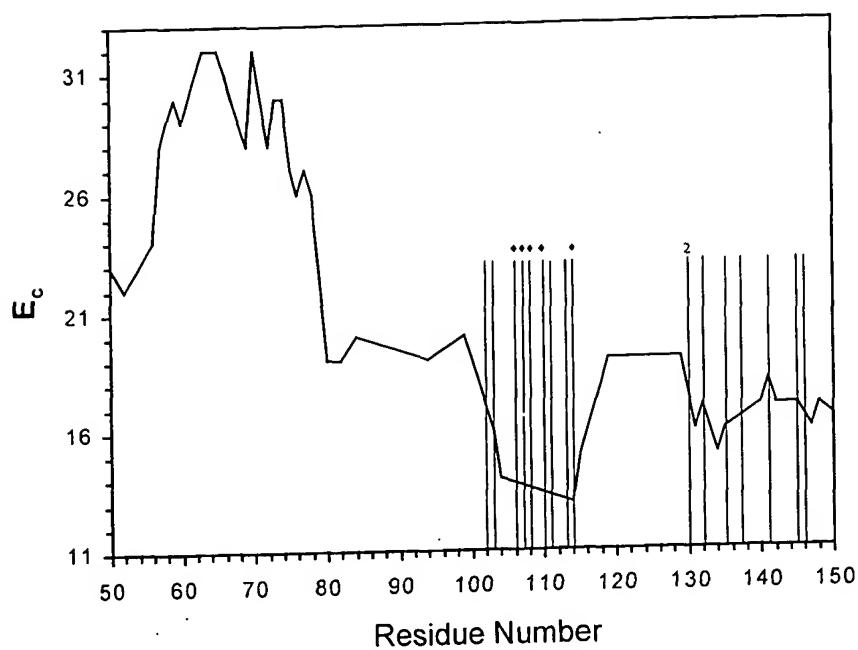


FIG. 5

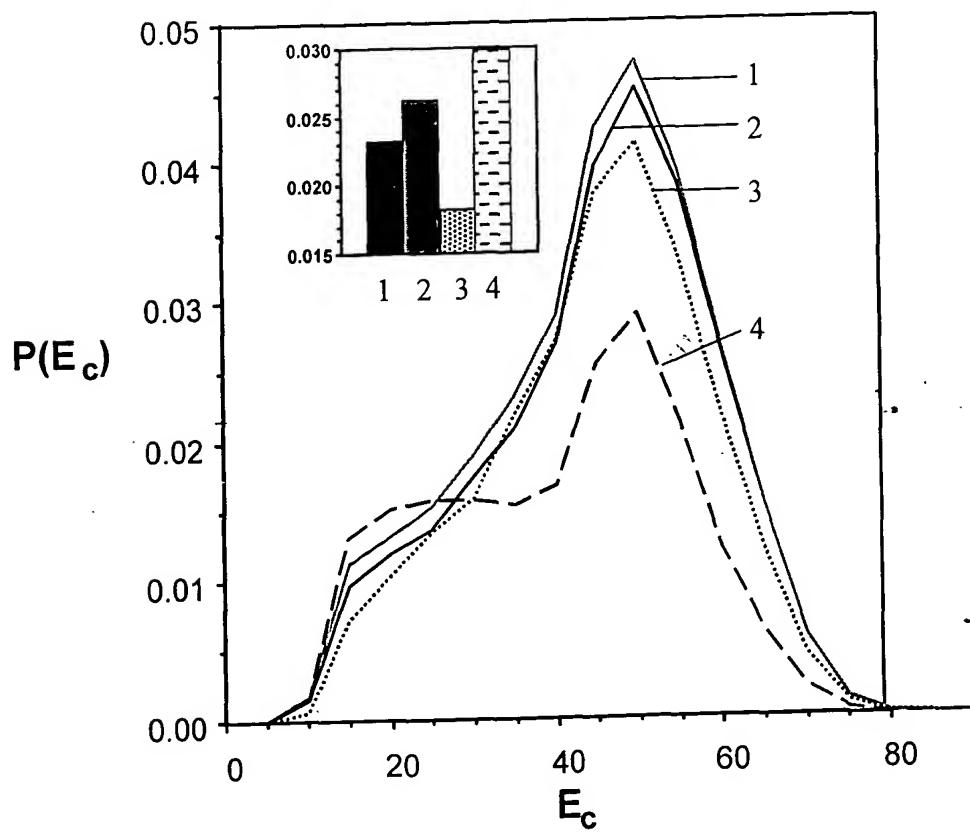
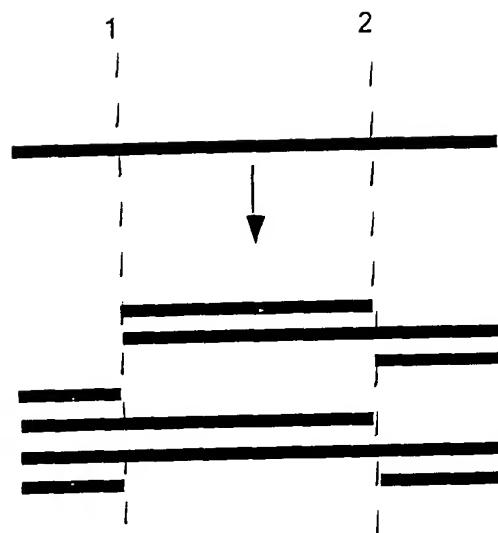


FIG. 6

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(A)

All possible recombinants
prepared by crossover
at positions 1 and 2



(B)

These can be prepared by
assembly of synthetic
fragments containing the
crossover positions

Requires fragments
(plus end primers):

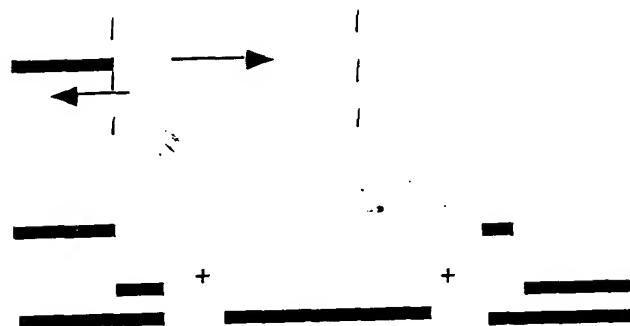
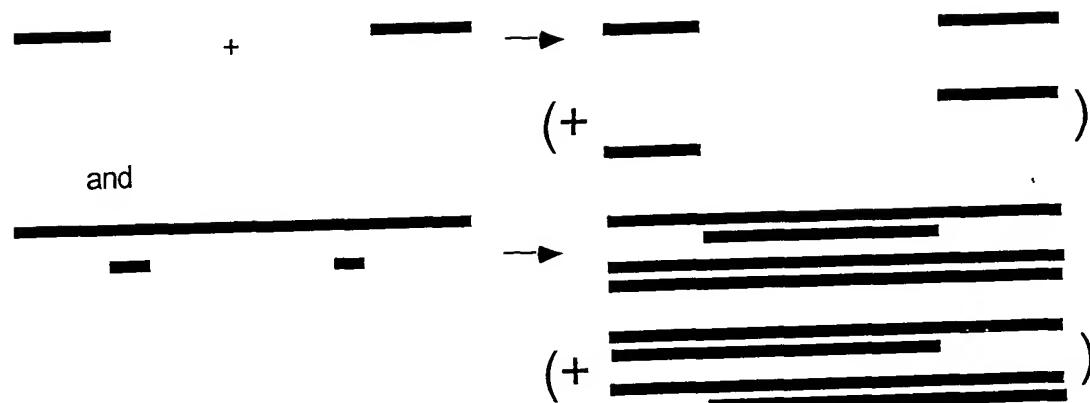


FIG. 7

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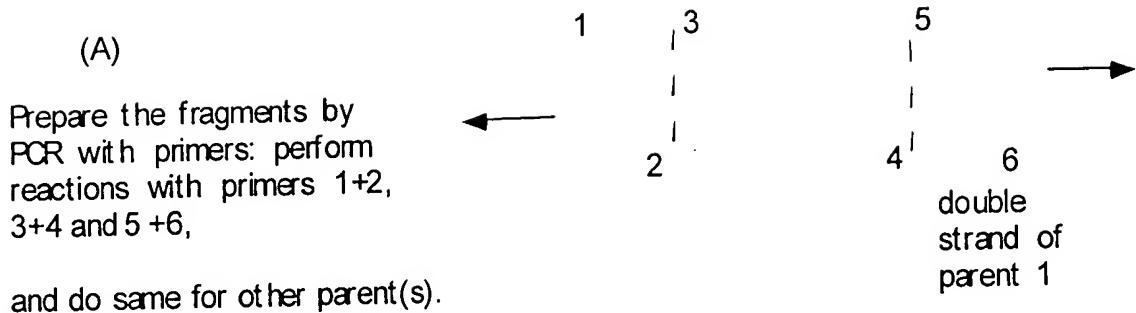
Extension of synthetic
fragments against a
parent template strand
and gap repair



heteroduplex recombination
(remove parent homoduplexes)

library of recombinants
with crossovers in regions
of non-identity

FIG. 8



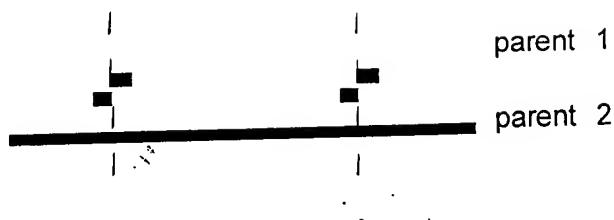
(B)

Reassemble fragments in a pool, by PCR with 1+ 6

FIG. 9

(A)

Prepare crossover primers designed to have crossovers at designated positions (2 primers for each position).



(B)

Fragment parent genes and PCR reassemble in the presence of the crossover primers to promote recombination at designated positions

FIG. 10

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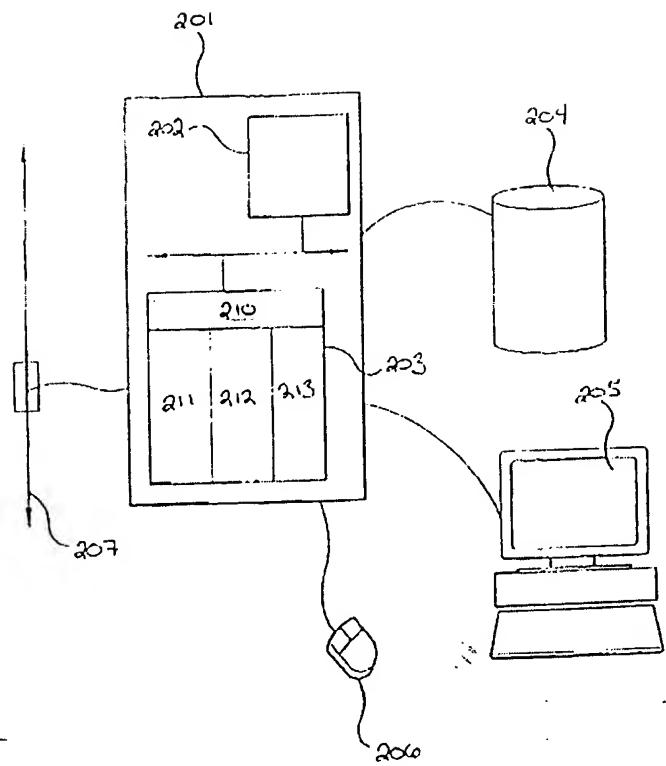
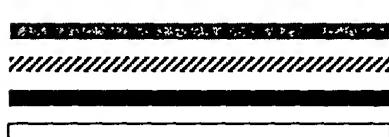


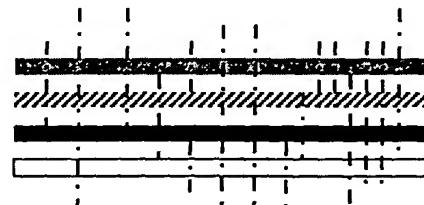
FIG. 11

Recombinant search algorithm

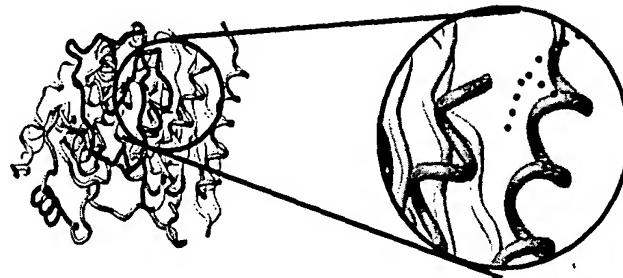
1. Align parent sequences with template structure



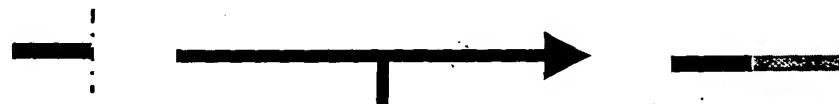
2. Determine all possible crossover points according to sequence identity algorithm



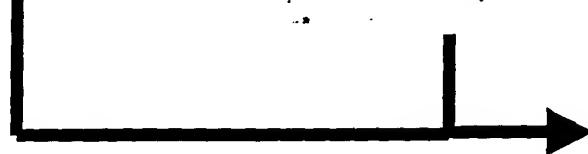
3. Calculate coupling matrix



4. Pick start parent at random and copy to offspring until a possible cut point is reached



5. Pick random number, if less than p , copy random new parent until next cut point is reached.



6. Determine crossover disruption of offspring gene

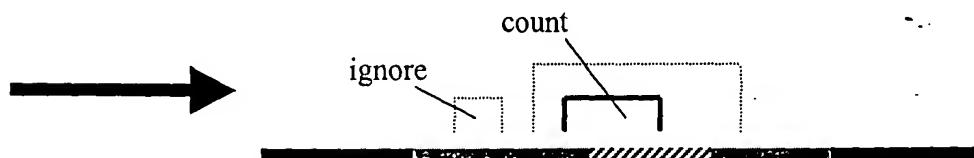


FIG. 12

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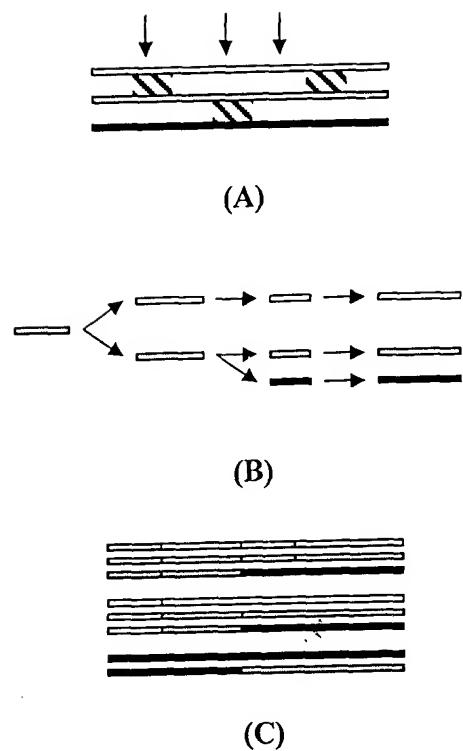


FIG. 13

DIRECTED EVOLUTION ALGORITHM

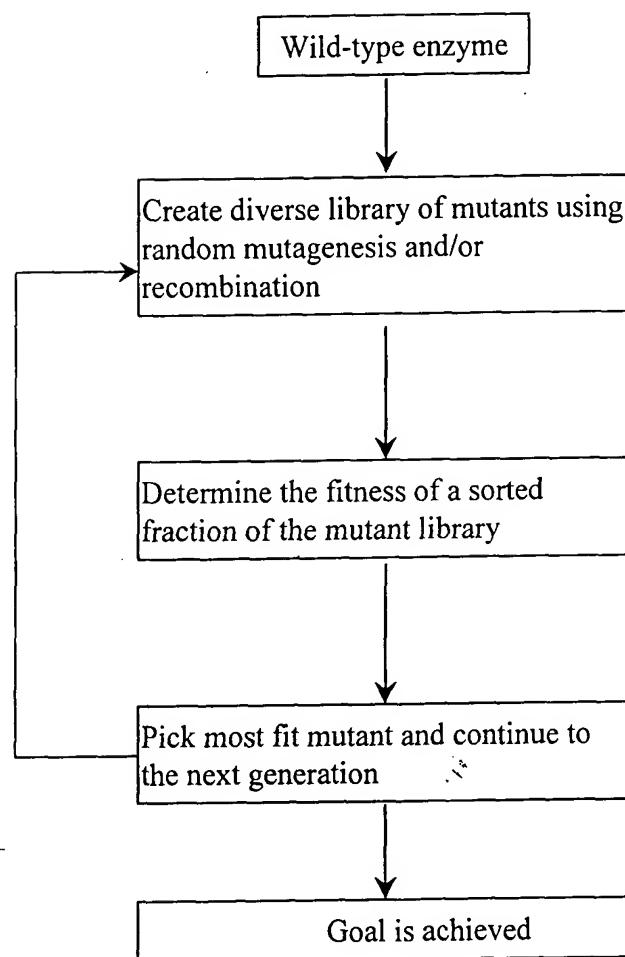


FIG. 14

F D E D F E D E D F E D

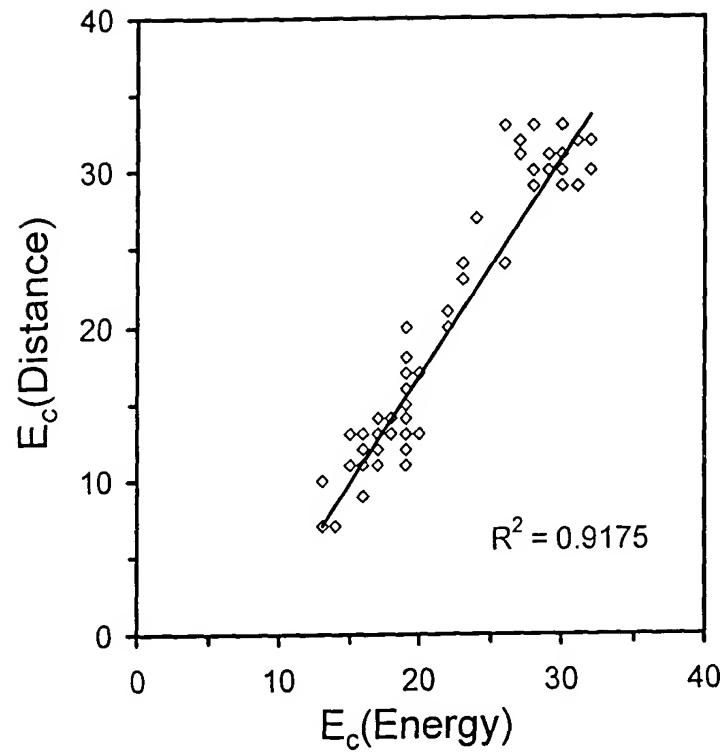


FIG. 15

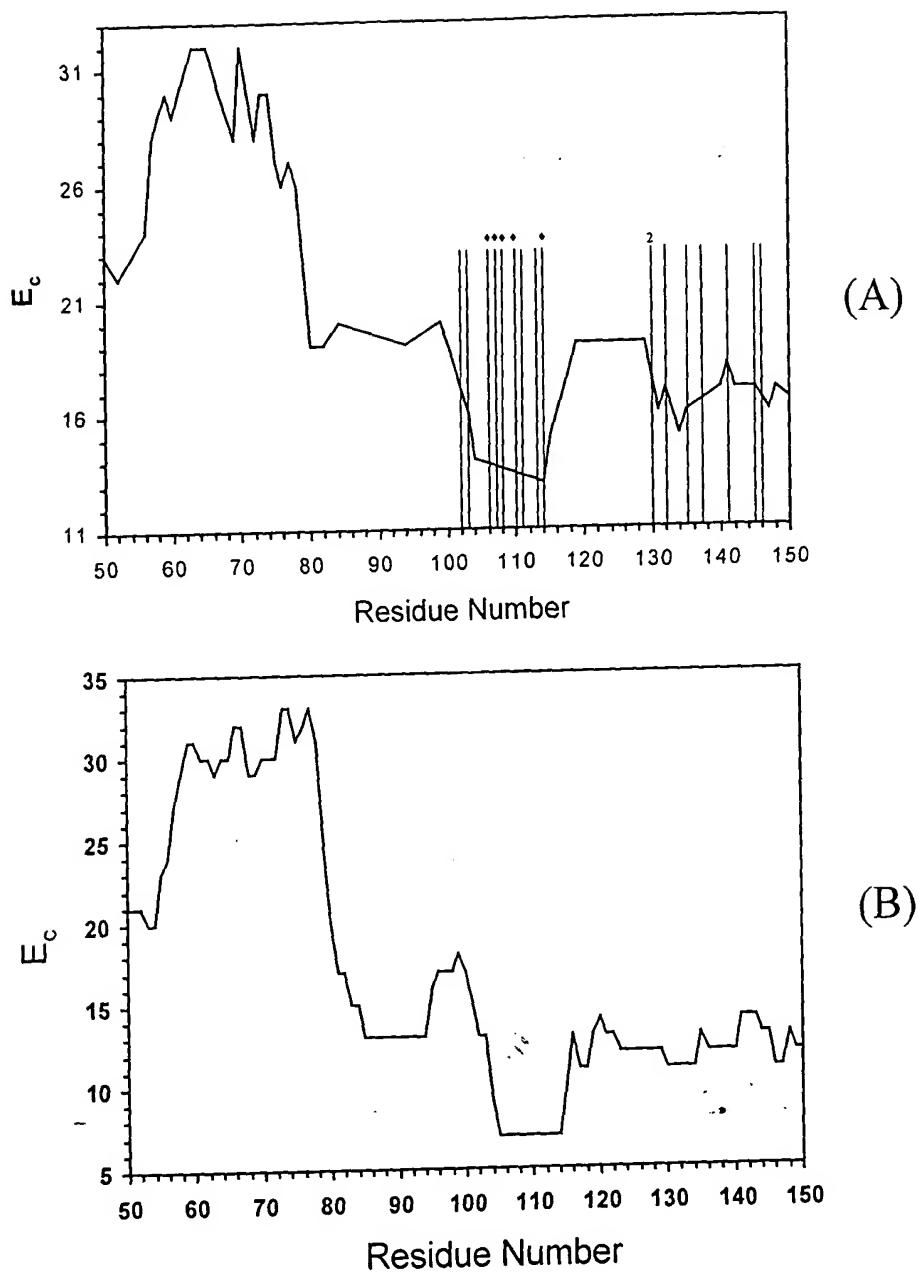
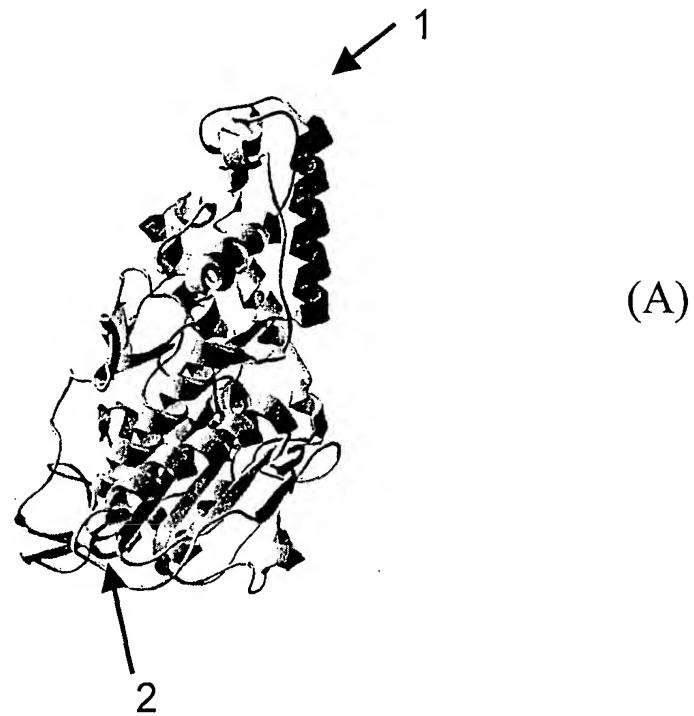


FIG. 16

FOOTNOTE: 89937003



Experimental Data:

| | wt | wt-insert | 1 | 2 |
|------------------|------|-----------|------|------|
| Tm(dC) | 52 | 55.2 | n.d. | 54.3 |
| Tm(dC) | 49.5 | 53.3 | 44.5 | 52.5 |
| t _{1/2} | 12.1 | 2586 | - | 87.5 |
| t _{1/2} | 53 | 138 | 4 | 308 |

(B)

Calculations:

| | All schema | | Fragments | | Z-score | |
|-----|------------|-------|-----------|-------|---------|--------|
| | av | stdev | 1 | 2 | 1 | 2 |
| Ec | 19.260 | 4.090 | 10.770 | 8.124 | -2.076 | -2.723 |
| Ec* | 0.006 | 0.002 | 0.014 | 0.005 | 4.838 | -0.857 |

FIG. 17

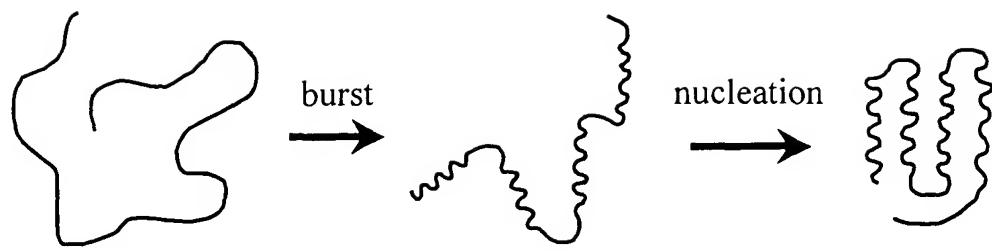


FIG. 18

The contact map shows residues that are distant (black) and residues that are close (white). If a given segment, , folds an above average number of residues into a given sphere size, then it is compact.

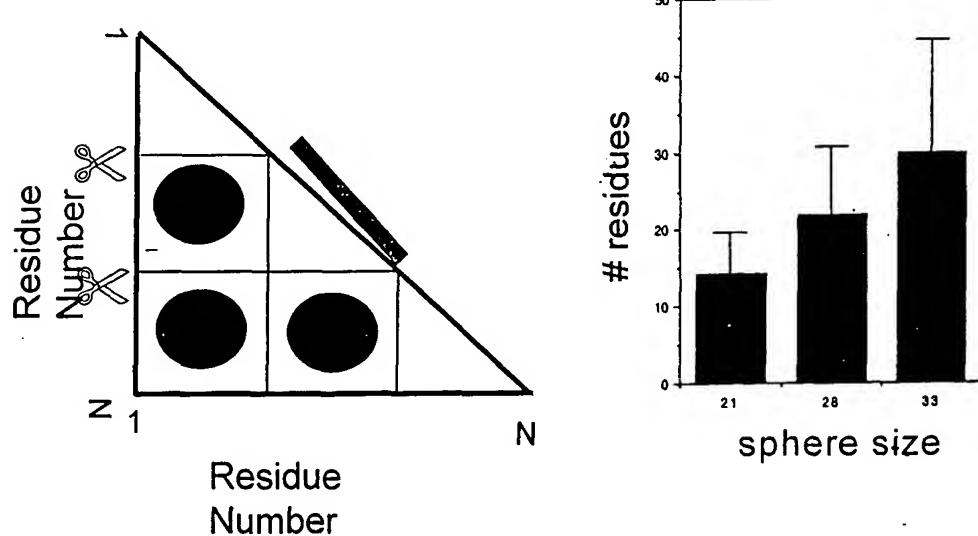
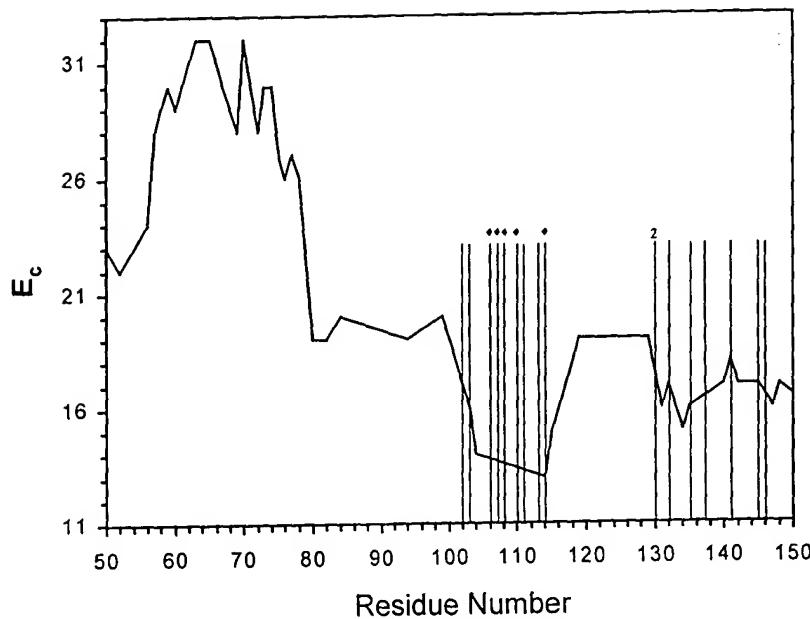
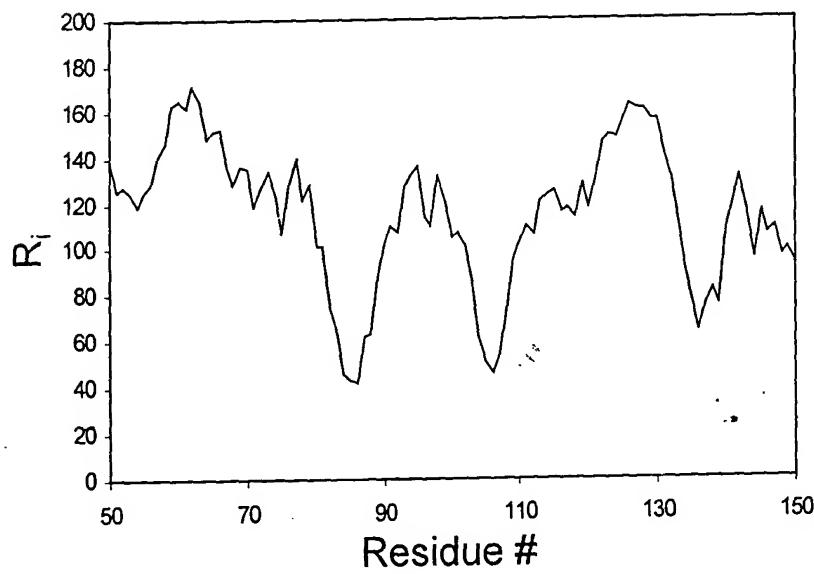


FIG. 19

200 190 180 170 160 150 140 130 120 110



(A)



(B)

FIG. 20

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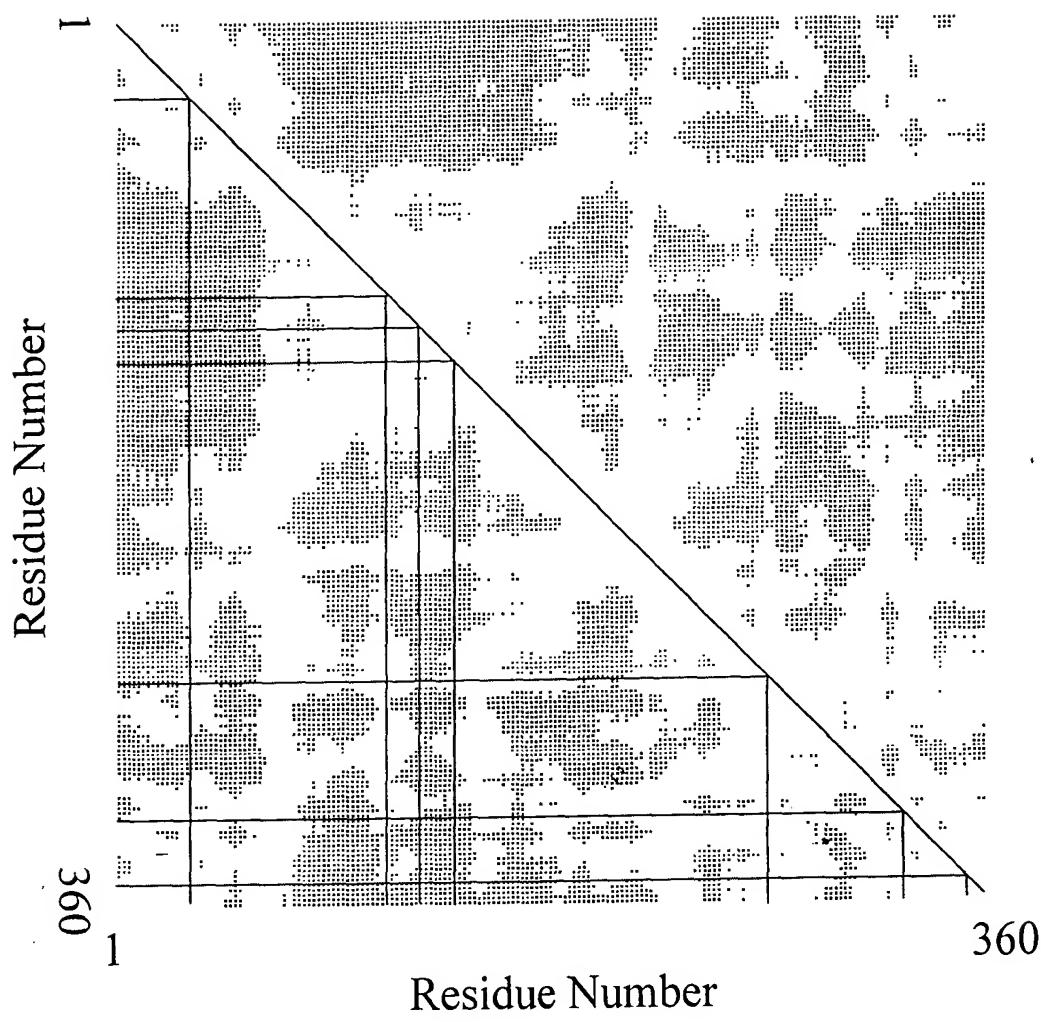


FIG. 21

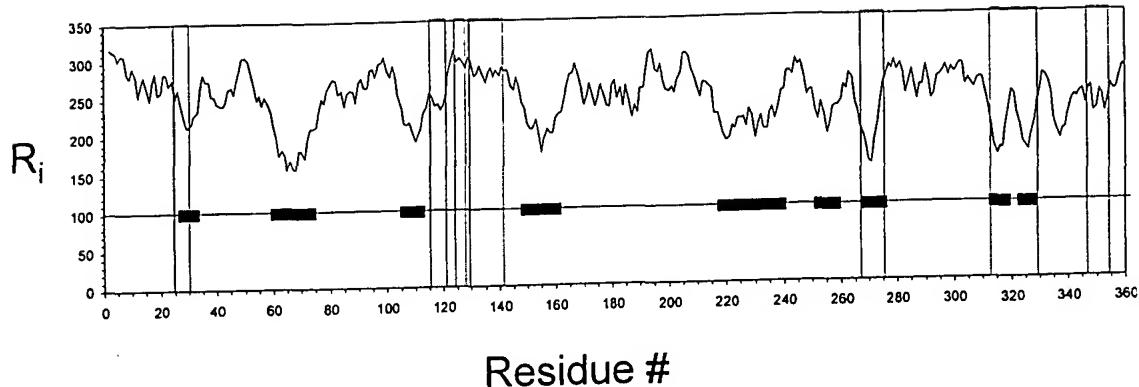
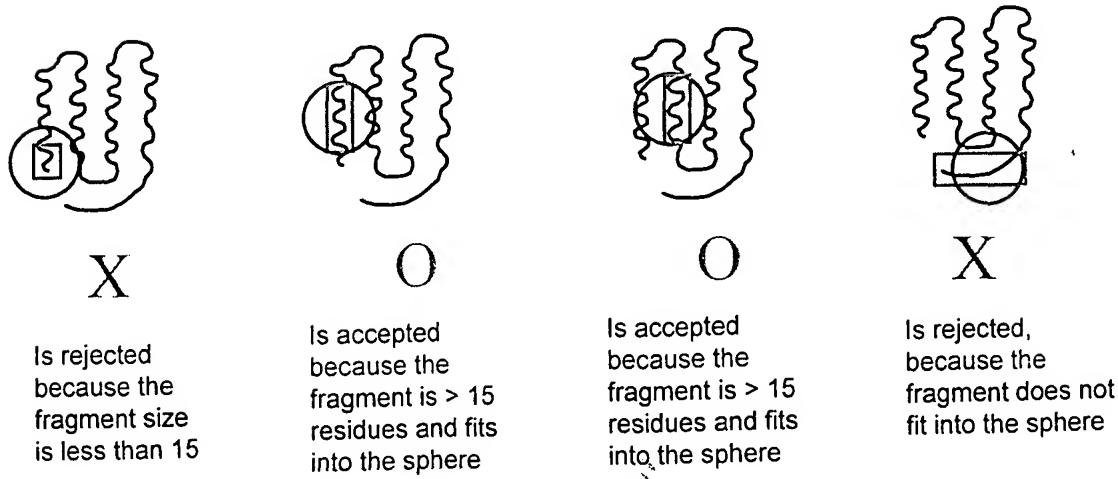


FIG. 22



- (1) Pick a sphere size (21 angstroms, like Go-Gilbert) and a disruption threshold; (2) Scan protein using segments at least the average number of residues for that sphere size or greater (e.g., >15 for 21 angstrom sphere); 3) Check the disruption of all the compact fragments identified in step 2. If the fragment has a disruption above a threshold value, keep it; otherwise, throw it out; 4) If the compact unit is disruptive, increment the schema disruption measure for all of the residues in the fragment by one. This indicates that crossovers within the fragment are disfavored.

FIG. 23

1 0 0 1 1 6 6 5 6 1 0 2 5 0 1

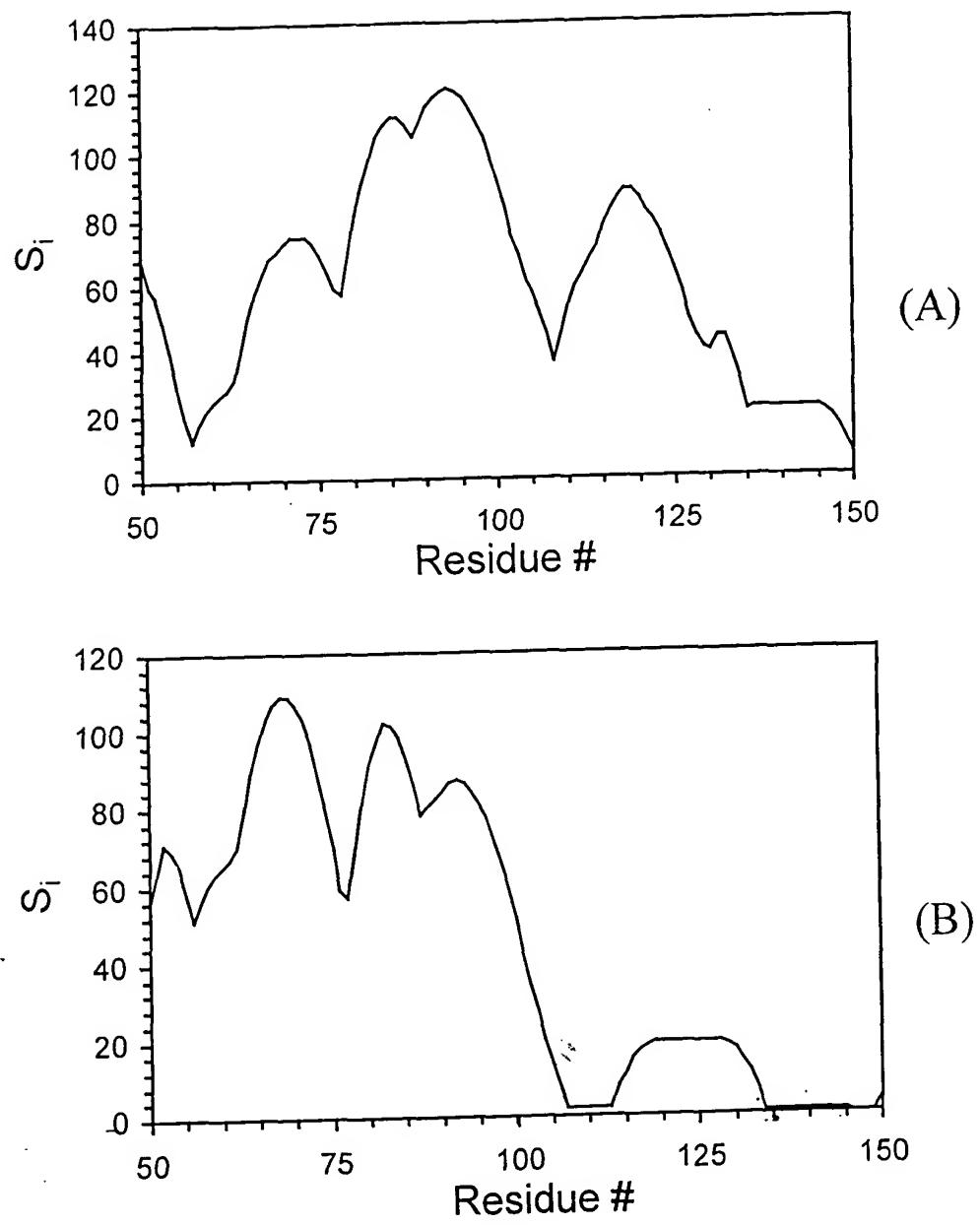


FIG. 24

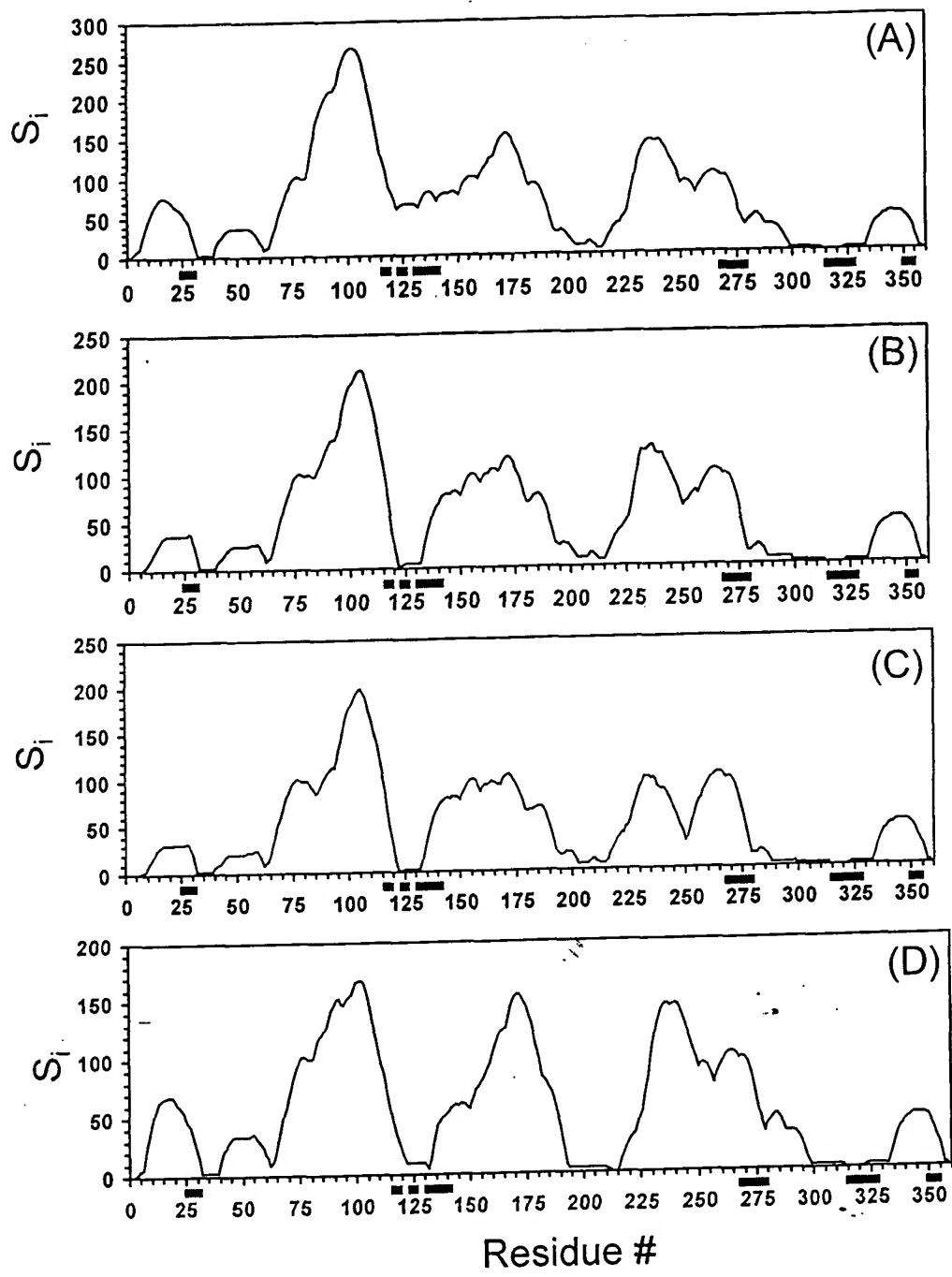


FIG. 25

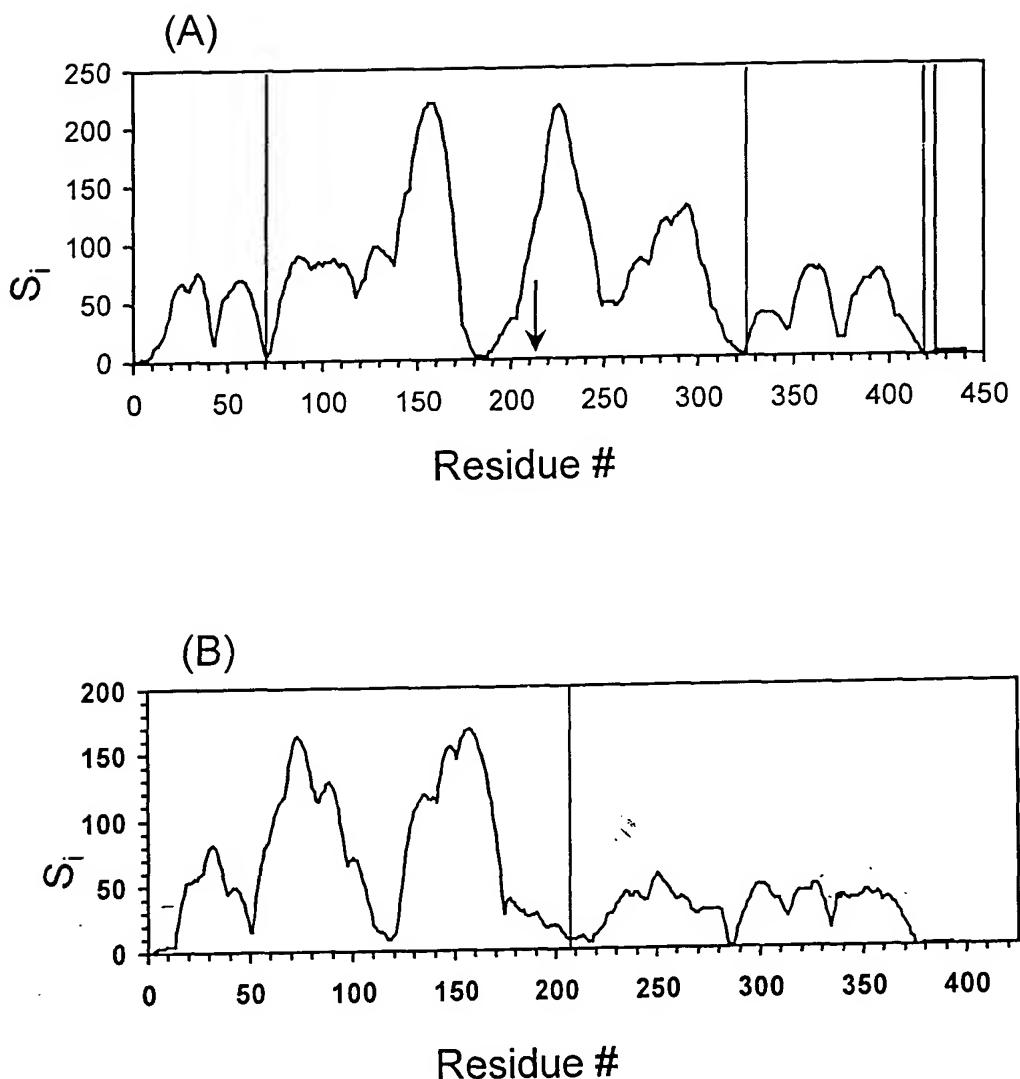


FIG. 26

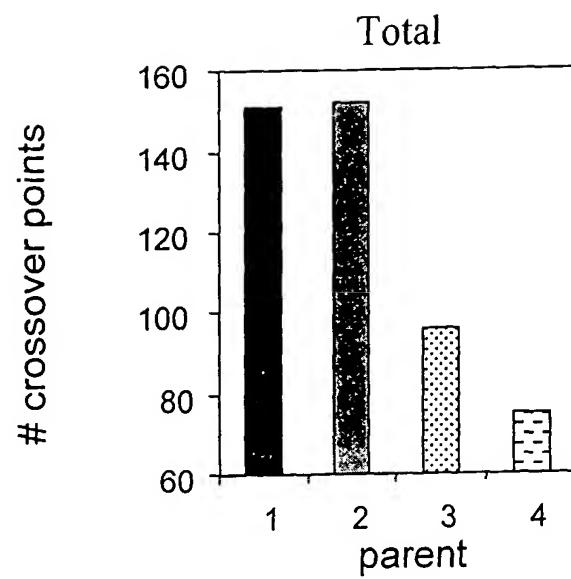


FIG. 27A

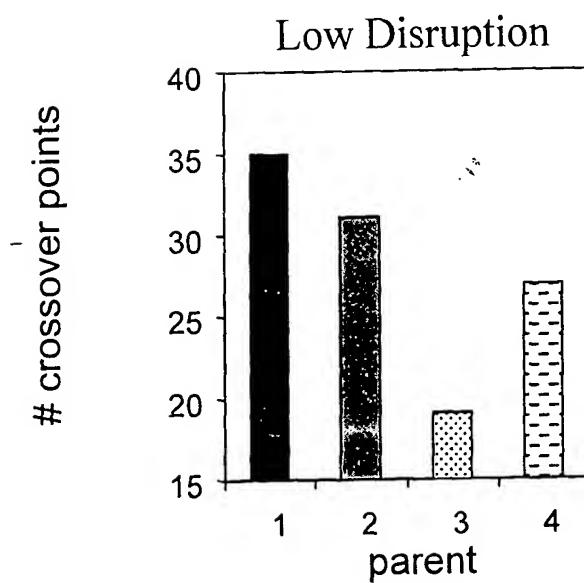


FIG. 27B

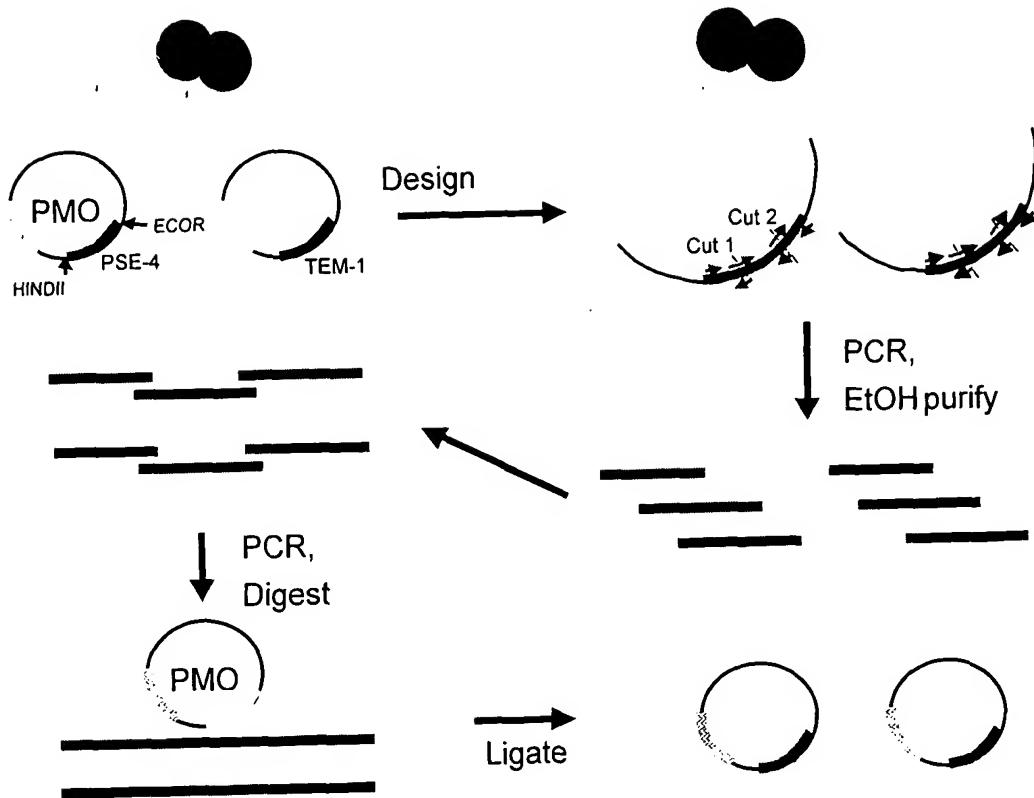


FIG. 28

Schema Disruption

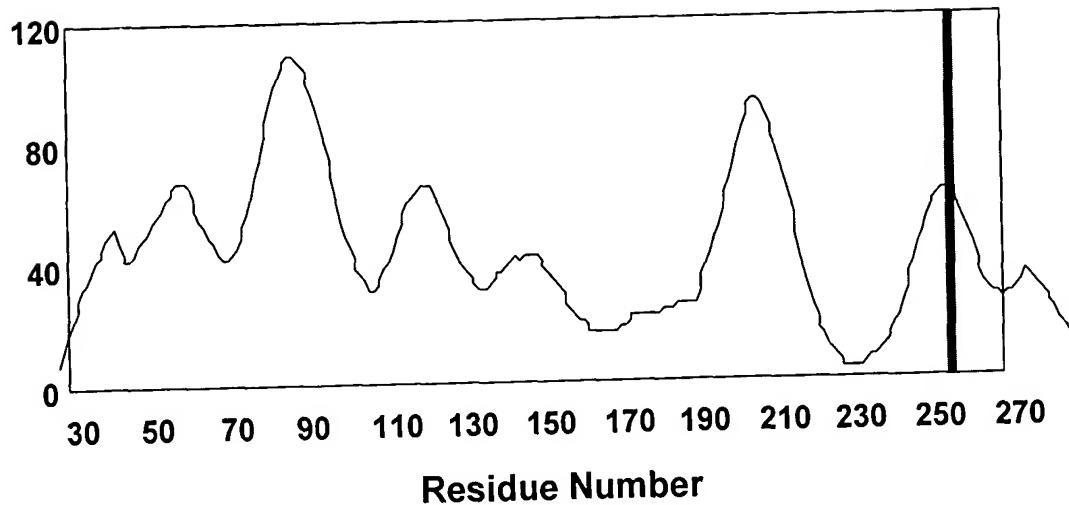


FIG. 29

Schema Disruption

4.00 3.96 3.92 3.88 3.84 3.80 3.76 3.72 3.68 3.64 3.60 3.56 3.52 3.48 3.44 3.40 3.36 3.32 3.28 3.24 3.20 3.16 3.12 3.08 3.04 3.00 2.96 2.92 2.88 2.84 2.80 2.76 2.72 2.68 2.64 2.60 2.56 2.52 2.48 2.44 2.40 2.36 2.32 2.28 2.24 2.20 2.16 2.12 2.08 2.04 2.00 1.96 1.92 1.88 1.84 1.80 1.76 1.72 1.68 1.64 1.60 1.56 1.52 1.48 1.44 1.40 1.36 1.32 1.28 1.24 1.20 1.16 1.12 1.08 1.04 1.00 0.96 0.92 0.88 0.84 0.80 0.76 0.72 0.68 0.64 0.60 0.56 0.52 0.48 0.44 0.40 0.36 0.32 0.28 0.24 0.20 0.16 0.12 0.08 0.04 0.00

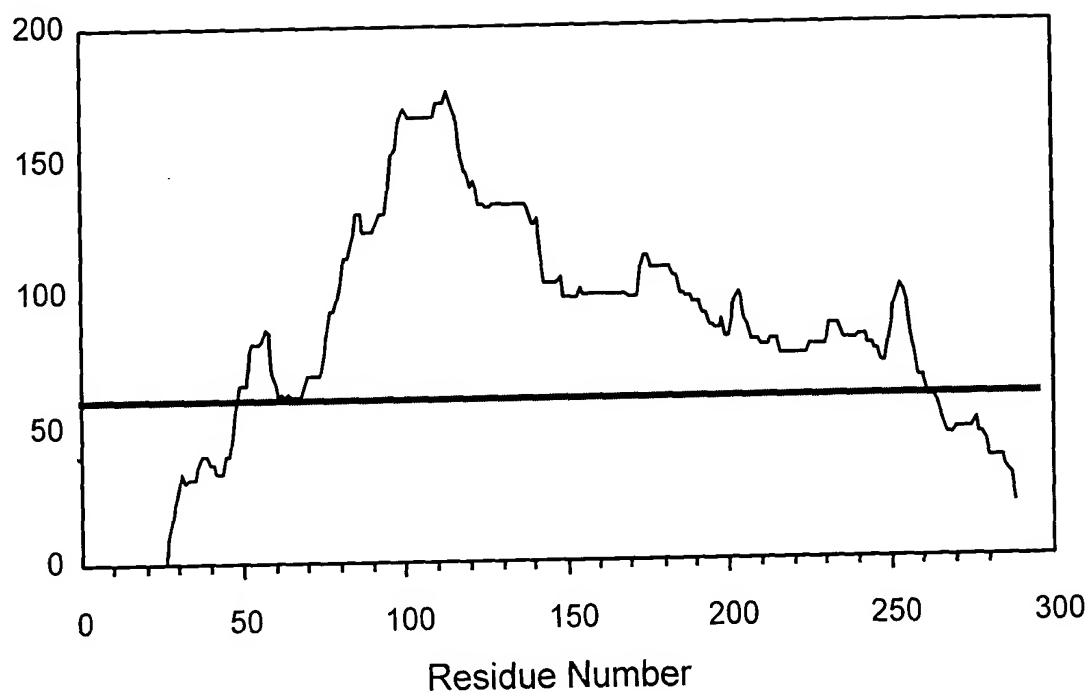


FIG. 30

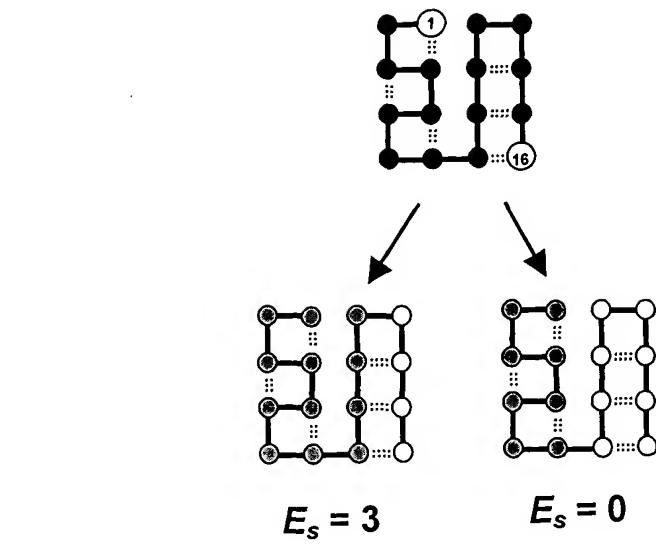


FIG. 31A

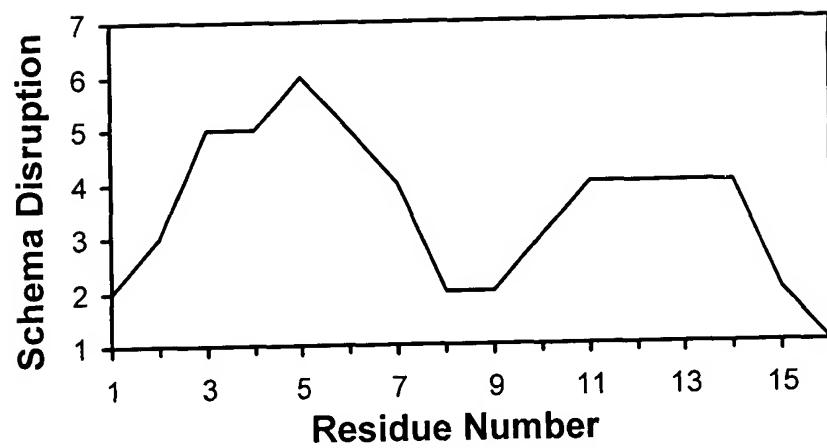


FIG. 31B

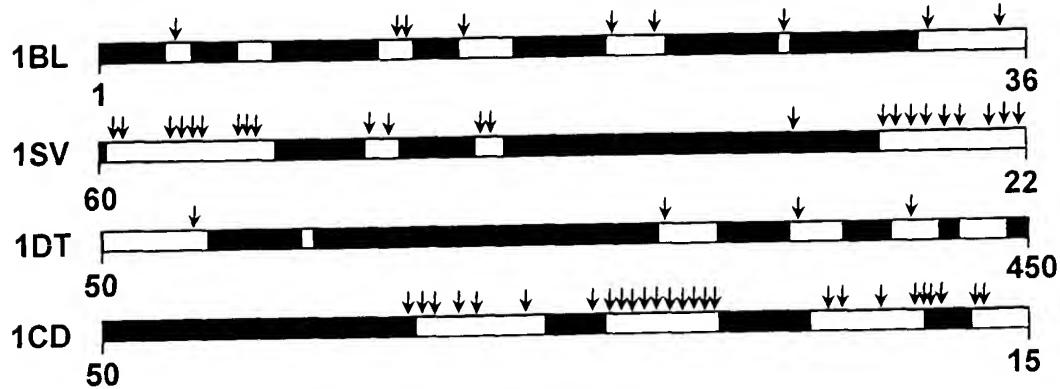


FIG. 32

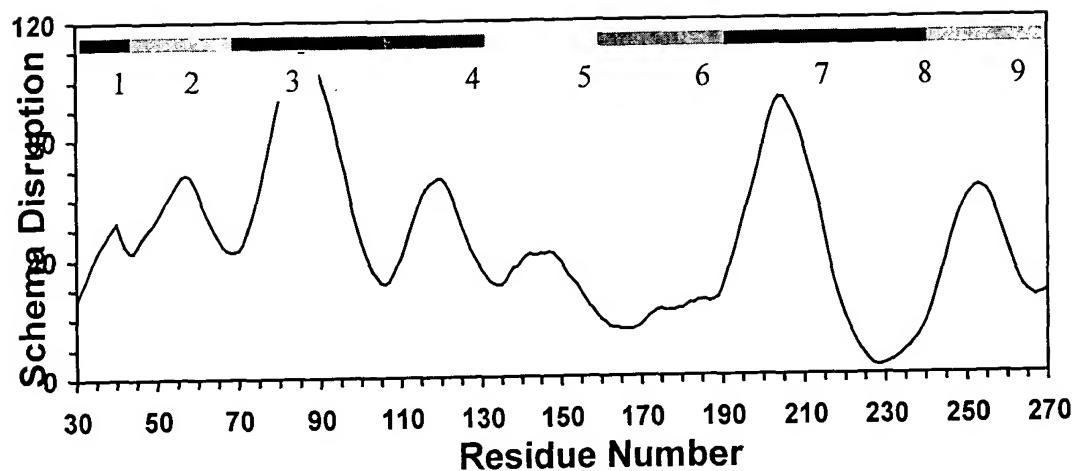


FIG. 33

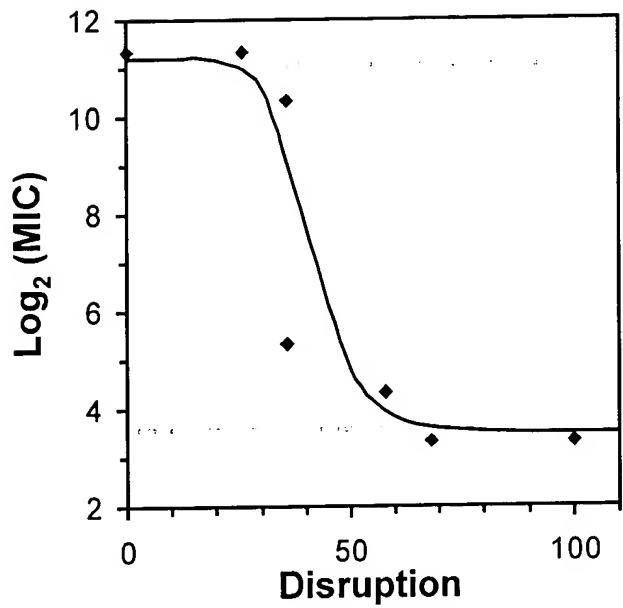


FIG. 34